Mutations of genes in synthesis of the carotenoid precursors of ABA lead to pre-harvest sprouting and photo-oxidation in rice

Jun Fang1,2,†, Chenglin Chai1,2,†, Qian Qian3,†, Chunlai Li1,2, Jiuyou Tang1,2, Lei Sun4, Zejun Huang1,2, Xiaoli Guo1,2, Changhui Sun1, Min Liu1, Yan Zhang5, Qingtao Lu5, Yiqin Wang1, Congming Lu5, Bin Han6, Fan Chen7, Zhukuan Cheng1 and Chengcai Chu1,*

1State Key Laboratory of Plant Genomics and National Centre for Plant Gene Research (Beijing), Institute of Genetics and Developmental Biology, Chinese Academy of Sciences (CAS), Beijing 100101, China,
2Graduate University of the CAS, Beijing 100039, China,
3State Key Laboratory of Rice Biology, China National Rice Research Institute, Chinese Academy of Agricultural Sciences, Hangzhou 310006, China,
4Centre for Biological Electron Microscopy, Institute of Biophysics, CAS, Beijing 100101, China,
5State Key Laboratory of Photosynthesis and Environmental Molecular Physiology, Institute of Botany, CAS, Beijing 100093, China,
6National Centre for Gene Research, CAS, Shanghai 200233, China, and
7Laboratory of Molecular and Developmental Biology, Institute of Genetics and Developmental Biology, CAS, Beijing 100101, China

Received 26 November 2007; revised 28 December 2007; accepted 3 January 2008.
*For correspondence (fax +8610 6487 7570; e-mail ccchu@genetics.ac.cn).
†These authors contributed equally to this article.

OnlineOpen: This article is available free online at www.blackwell-synergy.com

Summary

Pre-harvest sprouting (PHS) or vivipary in cereals is an important agronomic trait that results in significant economic loss. A considerable number of mutations that cause PHS have been identified in several species. However, relatively few viviparous mutants in rice (Oryza sativa L.) have been reported. To explore the mechanism of PHS in rice, we carried out an extensive genetic screening and identified 12 PHS mutants (phs). Based on their phenotypes, these phs mutants were classified into three groups. Here we characterize in detail one of these groups, which contains mutations in genes encoding major enzymes of the carotenoid biosynthesis pathway, including phytoene desaturase (OsPDS), ß-carotene desaturase (OsZDS), carotenoid isomerase (OsCRTISO) and lycopene ß-cyclase (ß-OsLCY), which are essential for the biosynthesis of carotenoid precursors of ABA. As expected, the amount of ABA was reduced in all four phs mutants compared with that in the wild type. Chlorophyll fluorescence analysis revealed the occurrence of photoinhibition in the photosystem and decreased capacity for eliminating excess energy by thermal dissipation. The greatly increased activities of reactive oxygen species (ROS) scavenging enzymes, and reduced photosystem (PS) II core proteins CP43, CP47 and D1 in leaves of the Oscrtsol/phs3-1 mutant and OsLCY RNAi transgenic rice indicated that photo-oxidative damage occurred in PS II, consistent with the accumulation of ROS in these plants. These results suggest that the impairment of carotenoid biosynthesis causes photo-oxidation and ABA-deficiency phenotypes, of which the latter is a major factor controlling the PHS trait in rice.

Keywords: rice, pre-harvest sprouting, photo-oxidation, carotenoid biosynthesis, abscisic acid.

Introduction

The phenomenon of germination of cereal grains in the ear or panicle, usually under wet conditions shortly before harvest, is termed pre-harvest sprouting (PHS) or vivipary. Pre-harvest sprouting of cereal grains not only causes reduction of grain yield but also affects the quality of the grain. In Southeast Asia, PHS frequently occurs in rice due to
the long spell of rainy weather in early summer and autumn (Wan et al., 2006). In south China alone, heavy PHS sometimes occurs in >6% of the rice acreage, which could be up to 20% for hybrid rice (Guo et al., 2004).

Previous work demonstrated that mutants impaired in abscisic acid (ABA) biosynthesis or responsiveness, such as maize viviparous (vp), Arabidopsis ABA-deficient (aba) and ABA-insensitive (abi) mutants, often produce precociously germinating seeds (McCarty, 1995). At least 10 viviparous mutants have been identified from maize (Zea mays L.), most of which (vp2, vp5, vp7, vp9, w3, y3, and y9) were blocked in biosynthesis of the carotenoid precursors for de novo ABA synthesis (Figure S1; Singh et al., 2003). In the early steps of carotenoid biosynthesis, the head-to-head condensation of C20 geranylgeranyl diphosphate (GGPP) molecules to produce a C40 carotenoid phytoene (colorless) is mediated by a soluble enzyme called phytoene synthase (PSY), which is the first committed step in carotenoid synthesis. Subsequently the phytoene undergoes four desaturation reactions with the production of lycopene (Cunningham and Gantt, 1998). Then, a series of steps including cyclization and hydroxylation reactions take place to yield α-carotene, β-carotene, lutein, xanthophyll, and zeaxanthin. The C40 carotenoid precursor is cleaved and followed by a two-step conversion of the intermediate xanthoxin to ABA via ABA aldehyde (Schwartz et al., 2003; Xiong and Zhu, 2003).

The maize Vps5 gene was found to encode a phytoene desaturase (PDS), and transgenic rice plants harboring the PDS-RNAi construct showed a clear albino phenotype (Hable et al., 1998; Miki and Shimamoto, 2004). The maize vp9 mutant and the non dormant-1 (nd-1) mutant of sunflower (Helianthus annuus L.) have mutations in the gene coding for zeta-carotene desaturase (ZDS; Conti et al., 2004; Matthews et al., 2003). The carotenoid isomerase (CRTISO) gene was cloned from the tangerine mutant of tomato and ccr2 mutant of Arabidopsis (Isaacson et al., 2002; Park et al., 2002). The Vp7/Ps7 gene encodes a lycopene-β-cyclase and is necessary for the accumulation of both ABA and carotenoid zeaxanthin in mature maize embryos; the mutant is easily discernible as it has pink kernels because of lycopene accumulation (Singh et al., 2003). These mutants containing defects in carotenoid precursor synthesis exhibit pleiotropic phenotypes, such as albino or pale green, non-viable seedlings and vivipary, due to deficiencies of carotenoid and ABA.

Within the thylakoid membranes of chloroplasts, carotenoids are found to be bound to specific protein complexes of photosystem I (PS I) and photosystem II (PS II), where they augment the light-harvesting capacity by absorbing light in the blue–green range of the visible spectrum (450–550 nm) and transferring the energy to chlorophylls (Holt et al., 2005; Moise et al., 2005). Carotenoid deficiency often causes aberrations in plastid ultrastructure, such as etioplasts from the cotyledon of dark-grown ccr2 mutants (carotenoid isomerase, CRTISO mutation) which lack prolamellar bodies (PLBs; Park et al., 2002). In addition, carotenoids play an essential role in photoprotection in plants. During photosynthesis, the excess absorbed energy can be eliminated as heat by de-excit ing 1Chl through the process of non-photochemical quenching of chlorophyll fluorescence (NPQ), minimizing the generation of harmful reactive oxygen species (ROS; Ma et al., 2003; Niyogi, 1999; Tracewell et al., 2001).

Besides, as accessory pigments in photosynthesis and photoprotectors preventing photo-oxidative damage, carotenoids can also be the precursors to the hormone ABA. Analysis of mutant and transgenic plants has provided strong evidence that ABA biosynthesis and responses to this phytohormone are clearly involved in the onset and maintenance of dormancy and inhibition of PHS (Bewley, 1997). Abscisic acid biosynthetic mutants fail to induce seed dormancy and exhibit a vegetative wilt phenotype. These phenotypes have been used to define and prove the function of ABA in seed dormancy and water relations. The seeds of ABA-deficient mutants of tomato (Solanum lycopersicum L., e.g. sitiens), Arabidopsis (e.g. aba) and maize treated with fluridone (a carotenoid biosynthesis inhibitor) germinate readily when placed in water and sometimes show vivipary (Fong et al., 1983; Groot and Karssen, 1992; Leon-Kloosterziel et al., 1996a).

Another phytohormone, gibberellin (GA), is also involved in the release from dormancy of various species. It has been shown that GA-deficient mutants of Arabidopsis and tomato are dependent on exogenous GA for germination (Koornneef et al., 2002). Further work in maize showed that inhibition of GA biosynthesis mimics the effect of exogenous ABA in suppressing vivipary (White and Rivin, 2000; White et al., 2000). These results suggested that the ABA/GA ratio and not the absolute hormone content controls germination (Finch-Savage and Leubner-Metzger, 2006).

In contrast to the intensive molecular and genetic studies of seed dormancy in maize and Arabidopsis, the molecular mechanism of seed dormancy in rice is poorly understood, mainly because of the lack of available mutants with reduced dormancy. So far most research has focused on the quantitative trait locus (QTL) analysis of the natural differences in seed dormancy characteristics (Gonzalez-Guzman et al., 2004; Gu et al., 2004, 2006; Takeuchi et al., 2003; Wan et al., 2006). Only a few reports have been published on phs mutants and only one gene (OsABA1) has been cloned which was involved in ABA biosynthesis (Agrawal et al., 2001). We performed a large-scale screening of a rice T-DNA/Tos17 insertion mutant population to identify the genes involved in PHS. The warm and damp weather, a good elicitor of PHS, during the harvest season in Zhejiang Province on the southeast coast of China, enables us to isolate phs mutants efficiently. In an intensive screening of approximately 16 000 rice T1 mutant lines, we obtained 12
viviparous mutants. In this paper, four genes involved in carotenoid precursors of ABA biosynthesis were cloned. Our results suggested that the impairment in synthesis of the carotenoid precursors of ABA leads to photo-oxidation and PHS in rice, which will definitely be helpful for elucidating the molecular mechanisms of PHS in other crops such as wheat and barley that are susceptible to PHS.

Results

Identification and genetic analysis of the rice phs mutants

To identify rice phs mutants, we have screened a T-DNA/Tos17-mutagenized population (Nipponbare background) under field conditions in Hangzhou, downstream of the Yangtze River with a relatively high degree of humidity. Approximately 16 000 transgenic T1 lines were screened prior to harvest by visual inspection in the paddy field, and 27 putative mutants were identified with a viviparous phenotype. A representative mutant is shown in Figure S2(a). T2 seedlings were rescued, and five viviparous mutant lines (category I) exhibiting an enhanced wilty phenotype under conditions of water stress (category II; Figure S2d) or with embryo/seedling-lethal phenotypes (category III; Figure S2e). We present here a detailed characterization of six mutants which belong to category I (Table 1).

To further characterize these mutants the viviparous seedlings were rescued, and five viviparous mutant lines showed albino seedlings (Figure 1a), these homozygous plants eventually died at about 4 weeks after germination. Interestingly, the homozygous seeds of two mutant lines among them had pink embryos (Figure 1b). Moreover, one of the viviparous mutant lines, T09, developed alternating green and yellow crossbands on the leaf blades at the tillering stage (Figure 1c), like rice zebra mutants previously described (Kusumi et al., 2000), and finally showed pale green on entire plants when mature (Figure 1d).

Except for the T09 mutant, all the other viviparous mutants (T1 plants) are lethal due to lack of pigments, therefore only two genotypes – heterozygous and wild type in the T1 mutant seeds – could germinate after sowing. Statistical analysis of segregation ratios of viviparous and non-viviparous plants demonstrated that the segregation of viviparous and non-viviparous plants is in line with the expected 2:1 ratio (Table S2). Furthermore, in T2 progeny (seeds in the ear) obtained from the viviparous plants, the mutant phenotype segregated in 3:1 (Wt: vivipary, data not shown), indicating that each mutant phenotype was caused by a single recessive mutation. Subsequently genetic analysis suggested that these six mutants could be assigned to four loci and were consecutively designated as phs1 through phs4, respectively, of which two mutant alleles were identified in phs2 (phs2-1 and phs2-2) and phs4 (phs4-1 and phs4-2; see Table 1). In addition, we later identified another two phs3 alleles by the zebra phenotype (see below).

Molecular cloning of PHS1 through PHS4 genes

Genetic analysis suggested that T-DNA was not co-segregated with the mutant phenotype in all these mutants (data not shown). We then made a rough mapping to identify candidate genes in these mutants, and the PHS genes were located to different chromosomes (Table 1 and Figure S5).

Because only mutants specifically blocked in the carotenoid biosynthetic pathways can manifest as a combination of vivipary with albino phenotype (Wurtzel et al., 2001), we presumed that these mutants may carry lesions in key genes for biosynthesis of carotenoids or ABA. So the predicted amino acid sequences of ABA biosynthesis genes from Arabidopsis were used as probes to screen for biosynthesis of carotenoids or ABA. So the predicted amino acid sequences of ABA biosynthesis genes from Arabidopsis were used as probes to screen in silico all available rice databases. In these four regions where mutations phs1 through phs4 mapped, four candidate genes appearing to be orthologs of Arabidopsis genes were found in the carotenoid biosynthesis pathway (Table 1), including a PDS-like gene (phs1), a ZDS-like gene (phs2-1 and phs2-2), a carotenoid isomerase-like gene (phs3-1) and a β-LCY-like gene (phs4-1 and phs4-2). To examine whether these candidate genes carried mutations in the respective

Table 1 Rice pre-harvest sprouting mutants and corresponding genes

<table>
<thead>
<tr>
<th>Category</th>
<th>Original line</th>
<th>Gene</th>
<th>Arabidopsis homolog</th>
<th>Mutation</th>
<th>Chromosome location</th>
<th>Protein (a.a.)</th>
<th>Accession no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>T01</td>
<td>OsPDS</td>
<td>AT4G14210</td>
<td>Insertion</td>
<td>Chr.3, 20.3cM</td>
<td>566</td>
<td>LOC_Os030g08570</td>
</tr>
<tr>
<td></td>
<td>HF807</td>
<td>OsZDS</td>
<td>AT3G04870</td>
<td>Insertion</td>
<td>Chr.7, 41.7cM</td>
<td>578</td>
<td>AK065213</td>
</tr>
<tr>
<td></td>
<td>HG4123</td>
<td></td>
<td></td>
<td>Deletion</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>T09</td>
<td>OsCRTISO†</td>
<td>AT1G06820</td>
<td>Substitution</td>
<td>Chr.11, 84.6cM</td>
<td>586</td>
<td>EF417892</td>
</tr>
<tr>
<td></td>
<td>HC2621</td>
<td>β-OsLYC</td>
<td>AT3G10230</td>
<td>Deletion</td>
<td>Chr.2, 23.3cM</td>
<td>493</td>
<td>OC_Os02g09750</td>
</tr>
<tr>
<td></td>
<td>HD1449</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

†The nucleotide sequence of OsCRTISO reported in this paper has been submitted to GenBank under accession number EF417892.
In phs3-1, a G-to-C transition was identified at position 1314 bp starting from ATG, and the phs4-2 allele has a 25-bp deletion in 473–497 bp (the putative translation start codon is referred to as +1). The deletions caused a frame-shift and subsequently created a new stop codon that might result in immature translation (Figure 2g) and disrupt the normal function of the β-LCY gene. Expression analysis also showed that the β-OsLCY transcript was barely detectable in phs4 alleles compared with the wild type (Figure 2h).

To confirm the identity of these candidate genes, we also performed genetic complementation experiments. Since homozygous mutant seedlings from three genes (OsPDS, OsZDS, and β-OsLCY) are non-viable, the homozygous seeds were screened out simply from the heterozygous lines at an early stage of callus induction using mature embryos. The embryos of the homozygous mutant initiated an albino bud after 2 weeks’ culture on Murashige and Skoog medium (Murashige and Skoog, 1962) with addition of 2 mg l⁻¹ 2,4-dichlorophenoxyacetic acid (2,4-D) in the light, and their calli derived from homozygous seeds regenerated the albino shoot during later regeneration (Figure 3a). Genomic DNA fragments covering the respective 5’-upstream regions, entire genes and 3’-downstream regions of respective candidate genes were introduced into corresponding mutants by Agrobacterium-mediated transformation. In all cases, the mutant phenotypes were rescued when genomic fragments containing the candidate genes were introduced (Figure 3b,c, and Table S3). In contrast, the transgenic plants containing the empty vectors failed to rescue the phs mutants (data not shown).

Carotenoid profiles of phs mutants

To analyze carotenoid metabolism in the phs mutants, high-performance liquid chromatograph (HPLC) analysis was performed on the extracts prepared from light-grown wild-type and mutant seedlings. As shown in Figure 4a, carotenoids were almost undetectable in phs1 (Ospds mutants) and phs2-1 or phs2-2 (Oszds mutants) seedlings at 460 nm. However, lycopene accumulated in phs4-1 or phs4-2 (β-OsLCY mutants). Carotenoids, especially lutein, were dramatically reduced in light-grown phs3-1 (OsCRTISO mutants) and an all-trans lycopene precursor, prolycopene, was accumulated in dark-grown phs3-1 seedlings (data not shown). The Ospds mutant contained a peak at 287 nm with a retention time of 25.54 min, which was absent in the wild type (Figure 4b). The derived spectrum of the peak showed characteristics of

NG0489 from the Rice Tos17 Insertion Mutant database (http://tos.nias.affrc.go.jp/~miyao/pub/tos17/index.html.en) completely disrupts the function of CRTISO (Figure 2f). phs3-3 has a 30-bp deletion in exon 6. The deletions in phs3-1 and phs3-3 do not cause a frame-shift in the open reading frame (ORF).

The rice β-OsLCY gene has no intron, and sequence analysis of phs4-1 mutant showed a 1-bp deletion at the 1314 bp starting from ATG, and the phs4-2 allele has a 25-bp deletion in 473–497 bp (the putative translation start codon is referred to as +1). The deletions caused a frame-shift and subsequently created a new stop codon that might result in immature translation (Figure 2g) and disrupt the normal function of the β-LCY gene. Expression analysis also showed that the β-OsLCY transcript was barely detectable in phs4 alleles compared with the wild type (Figure 2h).

To confirm the identity of these candidate genes, we also performed genetic complementation experiments. Since homozygous mutant seedlings from three genes (OsPDS, OsZDS, and β-OsLCY) are non-viable, the homozygous seeds were screened out simply from the heterozygous lines at an early stage of callus induction using mature embryos. The embryos of the homozygous mutant initiated an albino bud after 2 weeks’ culture on Murashige and Skoog medium (Murashige and Skoog, 1962) with addition of 2 mg l⁻¹ 2,4-dichlorophenoxyacetic acid (2,4-D) in the light, and their calli derived from homozygous seeds regenerated the albino shoot during later regeneration (Figure 3a). Genomic DNA fragments covering the respective 5’-upstream regions, entire genes and 3’-downstream regions of respective candidate genes were introduced into corresponding mutants by Agrobacterium-mediated transformation. In all cases, the mutant phenotypes were rescued when genomic fragments containing the candidate genes were introduced (Figure 3b,c, and Table S3). In contrast, the transgenic plants containing the empty vectors failed to rescue the phs mutants (data not shown).

Carotenoid profiles of phs mutants

To analyze carotenoid metabolism in the phs mutants, high-performance liquid chromatograph (HPLC) analysis was performed on the extracts prepared from light-grown wild-type and mutant seedlings. As shown in Figure 4a, carotenoids were almost undetectable in phs1 (Ospds mutants) and phs2-1 or phs2-2 (Oszds mutants) seedlings at 460 nm. However, lycopene accumulated in phs4-1 or phs4-2 (β-OsLCY mutants). Carotenoids, especially lutein, were dramatically reduced in light-grown phs3-1 (OsCRTISO mutants) and an all-trans lycopene precursor, prolycopene, was accumulated in dark-grown phs3-1 seedlings (data not shown). The Ospds mutant contained a peak at 287 nm with a retention time of 25.54 min, which was absent in the wild type (Figure 4b). The derived spectrum of the peak showed characteristics of

© 2008 The Authors
phytoene with three main peaks (data not shown). In phs2 seedlings only small peaks of \( \gamma \)-carotene and its isomers were detected at 430 nm, which were absent in the wild type (Figure 4c).

These results were consistent with the mutated genes in the biosynthetic pathway of carotenoids, and carotenoid biosynthesis is indeed impaired in these mutants. The albino phenotypes of the phs mutants (\( \text{Ospds}, \text{Oszds}, \text{and} \beta\text{-OsLCy} \)) were due to chlorophyll and carotenoid deficiency. The pink embryo in phs4 suggested lycopene accumulation, and the phenotype of phs3 (\( \text{Os.crtiso} \) mutant) seemed to result from photobleaching.

**Cellular characters of the phs mutants**

Carotenoids are essential components of photosynthetic systems, which occur in thylakoid membranes in chloroplasts in association with chlorophyll–protein complexes (Green and Durnford, 1996; Moise et al., 2005). To assess the effect of the \( \text{PHS} \) mutations on chloroplast development, the leaves of 2-week-old seedlings of all \( \text{phs} \) mutants mentioned above were examined by transmission electron microscopy. In wild types, chloroplasts were well-developed and organized in the mesophyll cells (Figure 5a), in contrast, no plastid- or chloroplast-like structures were found in the mesophyll cells of albino mutants (\( \text{phs1}, \text{phs2}, \text{and} \text{phs4}; \) Figure 5b–d). In the mesophyll cells of \( \text{Os.crtiso} \) mutants, irregularly shaped and abnormally developed chloroplasts were observed, with reduced and irregularly organized thylakoid membranes (Figure 5e,f).

**OsLCY-RNAi plants and phs mutants suffered from photo-oxidative damages**

Due to the photoprotective function of carotenoids, plants deprived of these pigments suffered from photobleaching.
damage, especially under high light conditions (Niyogi, 1999; Sagar et al., 1988). Since the homozygous mutant seedlings from three genes (OsPDS, OsZDS, and β-OsLCY) are nonviable, we generated transgenic rice plants harboring the β-OsLCY-RNAi construct. It was demonstrated that β-OsLCY-RNAi plants and Oscr3ioso mutants showed a distinctly photobleached phenotype on leaves (Figure 6a,b), which was in accordance with the accumulation of ROS as revealed by nitro blue tetrazolium (NBT) staining (Figure 6c,d). Furthermore, compared with that in the wild type, higher activities of ROS-scavenging enzymes were found in both β-OsLCY-RNAi plant and phs3-1 mutant (Figure S3), suggesting that these two plants were under oxidative stress.

The maximal efficiency of PS II photochemistry ($F_v/F_m$) is often considered to be an indicator of PS II function (Demmig-Adams and Adams, 1992; Krause and Weis, 1991). Therefore, we further investigated PS II photochemistry in phs3-1/Oscr3ioso mutants and β-OsLCY-RNAi plants, as well as the wild type. The $F_v/F_m$ ratios in wild-type plants are above 0.8. However, phs3-1/Oscr3ioso mutant and β-OsLCY-RNAi plants show a decrease in the $F_v/F_m$ ratio, which is about 0.65 (Figure 7a,b).

The phs3-1/Oscr3ioso mutants and β-OsLCY-RNAi plants also show different responses of NPQ to increasing photosynthetic photon flux density (PPFD). When PPFD was <500 μmol m$^{-2}$ sec$^{-1}$, the phs3-1/Oscr3ioso mutants and β-OsLCY-RNAi plants showed almost the same value of NPQ in comparison with the wild type. With further increase in PPFD, NPQ increased more rapidly in wild-type plants than in the Oscr3ioso/phs3-1Mutants and β-OsLCY-RNAi plants, leading to much higher values of NPQ than for the latter two plants (Figure 7c,d).
To get further evidence for the photo-oxidative damage occurring in PS II, we examined the levels of essential PS II core proteins CP43, CP47 and D1 (Green and Durnford, 1996), which are the targets of ROS (Niyogi, 1999), in the leaves from the \textit{phs3-1} mutant and \textit{b-OsLCY}-RNAi plants, respectively. Western analysis showed that the level of these proteins in the \textit{phs3-1} mutant and \textit{b-OsLCY}-RNAi plants decreased to 64–77% of that in the wild type, indicating the occurrence of photo-oxidative damage in PS II core proteins (Figure 8).

**Figure 6.** Photobleaching phenotype (a, b) and the detection of ROS (c, d) of \textit{Oscrtiso} and \textit{b-LCY-RNAi} plants and the corresponding wild types.

**Figure 7.** The maximal efficiency of PS II photochemistry ($F_v/F_m$; a, b) and non-photochemical quenching of chlorophyll fluorescence (NPQ; c, d) of \textit{phs3-1} mutant and \textit{b-OsLCY-RNAi} plants. Standard deviations were obtained from six measurements.

ABA and GA synthesis are altered in \textit{phs} mutants

To investigate whether the reduced level of carotenoids in \textit{phs} mutants causes a decreased level of ABA we measured the ABA content in the respective \textit{phs} mutants by immunoassay. As shown in Figure 9a, the amount of ABA was reduced in all four mutants, but much more significantly in \textit{Ospsds}, \textit{Oszds}, and \textit{Os1cy} mutants compared with that in the wild type. Since ABA is also directly involved in the regulation of stomatal aperture, we examined the water loss characteristics of \textit{phs3-1/Oscrtiso} plants. Wild-type and mutant plants were grown in the soil under a normal irrigation regime. The water loss analysis was carried out at the tillering stage, and the results revealed a water loss of about 53% of fresh weight within 120 min in \textit{phs} mutants, whereas wild-type leaves showed only a 38% loss, indicating that the rate of water loss in \textit{phs} mutants was faster than in wild-type plants (Figure 9b).

To test whether \textit{phs} mutant can sense or respond to ABA, we cultured seeds [30 days after pollination (DAP)] from \textit{phs3-1} mutant and the wild type in water with or without ABA (of different concentrations) treatment (Figure S4). In all treatments, the \textit{phs3-1} mutant and the wild type grew with similar kinetics, indicating that the \textit{phs3-1} mutant has a normal response to ABA.

Abscisic acid and another important phytohormone, GA, which can directly antagonize ABA signaling during induction of dormancy, are two important regulators of seed dormancy. Therefore, we measured the ABA and GA content in homozygous \textit{phs4-1/b-Os1cy} seeds 30 DAP (\textit{phs4} homozygous seeds were easily discernible by their pink embryo). As expected, the ABA level in \textit{phs4-1/b-Os1cy} was lower than
that in the wild type; however, GA in phs4-1/β-OsLCY is higher than that in the wild type (Figure 10a). We further examined the expression of two ABA-regulated genes, rab16B and TRAB1, as well as a GA-induced gene, the α-amylase gene RAmy1A (AK073487), which were involved in embryo development and seed maturation (Hobo et al., 1999; Mitsui et al., 1996; O’Neill et al., 1990; Ono et al., 1996). The results demonstrated that the transcripts of rab16B and TRAB1 were significantly reduced in phs4-1/β-OsLCY seeds while the transcript of RAmy1A was increased (Figure 10b), suggesting that the ABA/GA ratio in phs4-1/β-OsLCY seeds is probably a switch for PHS.

**Discussion**

**Analysis of phs mutants in rice**

Under normal circumstances, germination in the ear of developing cereal grain is prevented by physiological mechanisms that include dormancy. Only under exceptional circumstances (e.g. rain late in grain maturation) may PHS occur. Since dormancy is a complex trait which is controlled by a large number of genes, not only with strong intergenic interactions but also strong interactions between genes and the environment. Thus, QTL analysis has often been the tool of choice for the dissection of dormancy and germination, and a large number of the QTLs controlling seed dormancy and germination have been identified in Arabidopsis, rice, wheat, and barley (Clerkx et al., 2004; Li et al., 2004; Prada et al., 2004). Comparative genomics approaches were also used to identify candidate gene(s) controlling dormancy and PHS based on the availability of the rice genome sequence, a large number of barley and wheat ESTs, and also the high co-linearity among rice, wheat, and barley (Li et al., 2004; Wilkinson et al., 2002).

Since PHS is a complex trait controlled by both environmental and genetic factors, we carried out intensive screening of the rice mutant population in south China, where long spells of rainy weather often occur in early summer and autumn and successfully isolated 27 rice phs mutants in the paddy fields.

In this study, we focused on phs mutants which belong to category I; using both map-based cloning and comparative genetics approaches, we have identified that the mutated genes in category I mutants encode major enzymes (OsPDS, OsZDS, OsCRTISO, and β-OsLCY) in the carotenoid biosynthesis pathway. The candidate genes were all confirmed via functional complementation. Further work with these rice phs mutants will provide novel insights into the role of these PHS genes in dormancy and germination.

**Carotenoid metabolism, plastid development and photo-oxidation in phs mutants**

In plant chloroplasts, carotenoids serve important roles in the photosynthetic apparatus as structural components and photosynthetic and photoprotective pigments (Green and Durnford, 1996; Niyogi, 1999; Niyogi et al., 2001). Just as expected, according to the enzymatic steps blocked, phs1 and phs2 accumulated phytoene and β-carotene in the light-
grown seedlings, respectively. The results are in accordance with maize vp5 and vp9 mutants with mutations in PDS and ZDS (Hable et al., 1998; Matthews et al., 2003). In case of \textit{phs4}, the mutation in lycopene \beta-cyclase leads to high lycopene accumulation in both seedlings and embryos from homozygous \textit{phs4} seeds. The accumulation of lycopene may be explained by the presumption that lycopene \epsilon-cyclase and lycopene \beta-cyclase constitute an enzyme complex in the thylakoid membranes of plant chloroplasts; in the absence of \beta-cyclase, the complex is destabilized, resulting in the loss of both \epsilon- and \beta-cyclase activity and so lycopene accumulates (Cunningham and Gantt, 1998). The results are consistent with 2-(4-chlorophenylthio)-triethylamine hydrochloride (CPTA)-treated barley seedlings (CPTA is a specific inhibitor of the cyclization of lycopene, especially lycopene \beta-cyclase; La Rocca et al., 2007). In addition, \textit{PSY} mRNA is virtually absent in rice endosperm, which does not provide the substrate for these downstream enzymes and is consequently unable to form downstream products (Schaub et al., 2005). Therefore, the \textit{phs4} mutant accumulates lycopene (showing a pink color) in the embryo but not in the endosperm.

Unlike \textit{phs1}, \textit{phs2}, and \textit{phs4} mutants, \textit{phs3} mutant show a non-lethal and ‘variegated’ phenotype. The mutation in the \textit{PHS3} gene results in a dramatic reduction of lutein in light-grown leaves and accumulation of protoplycopene in etiolated seedlings. This result was in agreement with other \textit{crtiso} mutants from tomato and \textit{Arabidopsis} (Isaacson et al., 2002; Park et al., 2002). Interestingly, the CRTISO activity could partially be substituted by light, i.e. photoisomerization, which may account for the survival of the \textit{phs3} mutant (Isaacson et al., 2002; Park et al., 2002).

Since carotenoids are located in the photosynthetic membrane in the form of chlorophyll–carotenoid–protein complexes and some carotenogenic enzymes are membrane-associated (Cunningham and Gantt, 1998; Green and Durnford, 1996), changes in carotenoid composition or the carotenogenic enzyme itself always lead to abnormal plastid development (Park et al., 2002; Welsch et al., 2000). It has been demonstrated that the location of \textit{PSY} protein influenced the formation of PLBs (Welsch et al., 2000), CRTISO is required in PS II assembly in cyanobacteria, and loss of CRTISO activity results in the absence of PLB in the \textit{ccr2} Arabidopsis mutant (Masamoto et al., 2004; Park et al., 2002). All the \textit{phs} mutants (\textit{phs1} through \textit{phs4}) showed dramatic changes in carotenoid composition and had abnormal chloroplasts or even none at all, suggesting the importance of these enzymes for plastid development.

Carotenoids could also act as antioxidants to quench excessive free radicals and ROS generated from photo-oxidation (Hirayama et al., 1994; Niyogi, 1999; Niyogi et al., 1998; Woodall et al., 1997). The increasing activities of ROS-scavenging enzymes in \beta-OsLCY RNAi plants and \textit{Ocrtiso} mutants suggested that these two plants suffered from oxidative damage. Non-photochemical quenching of chlorophyll fluorescence is one of the major indicators which are closely associated with the onset of harmless dissipation of the excess energy present in the pigment of light-harvesting complexes as heat (Gilmore, 1997; Niyogi, 1999). The significant decrease of NPQ in \beta-OsLCY RNAi plants and \textit{Ocrtiso} mutants indicated that excessive absorbed light could not be efficiently dissipated as heat and the excessive energy might result in ROS generation. Nitro blue tetrazolium staining further confirmed that considerable superoxide accumulated in both \beta-OsLCY RNAi plants and \textit{Ocrtiso} mutants. The value of \textit{Fv}/\textit{Fm} is normally in the range of 0.8–0.85 in healthy leaves independent of plant species, and a lower value indicated that a proportion of PS II reaction centers were damaged (Demig-Adams and Adams, 1992). The decreased \textit{Fv}/\textit{Fm} indicated that photoinhibition occurred in the \beta-OsLCY RNAi plants and \textit{Ocrtiso} mutants under normal conditions. Moreover, the decrease of PS II core proteins such as CP43, CP47, and D1 in \beta-OsLCY RNAi plants and \textit{Ocrtiso} mutants may provide further evidence for the occurrence of photo-oxidative damage in PS II. Taken together, these results further confirm that the investigated \textit{PHS} genes are involved in not only synthesis of the carotenoid precursors of ABA, but also chloroplast development and photoprotection. However, in the dark or in weak light the mechanism by which carotenoid deficiency influences plastid development needs to be further elucidated.

**ABA and pre-harvest sprouting in \textit{phs} mutants**

Seed dormancy and germination are regulated by a wide range of plant hormones, including ABA, ethylene, GA, and brassinosteroids, of which ABA is the primary mediator of seed dormancy (Koornneef et al., 2002). Abscisic acid-deficient or ABA-insensitive Arabidopsis mutants show reduced seed maturation and dormancy (Finch-Savage and Leubner-Metzger, 2006; Koornneef et al., 2002; Leon-Kloosterziel et al., 1996b). Unlike in Arabidopsis, in cereal plants such as maize embryos from ABA-deficient mutants germinate precociously on the ear. However, the absolute level of ABA in the seeds does not always perfectly correlate with the dormancy and germination events, suggesting that other modulating factors are also involved (White et al., 2000). It has been presumed that dormancy released by after-ripening and stratification caused a switch to ABA catabolism resulting in a decrease in ABA content in the embryo and a corresponding increase in inactive ABA metabolites (Gubler et al., 2005).

The amount of GA also plays an important role in controlling seed dormancy and germination. Previous studies revealed that paclobutrazol treatment of the ear suppresses vivipary in ABA-deficient seeds (White and Rivin, 2000).
2000). Here, we have demonstrated that the impaired biosynthesis of the carotenoid led to significant reduction of ABA content in the phs mutants. However, these mutants have a normal response to ABA. Interestingly, the amount of GA in phs4-1 seeds was significantly increased; the increased GA might result from a reduced flux of GGPP to carotenogenesis since GGPP is the common precursor for both GA and carotenoid biosynthesis (Rodriguez-Conception et al., 2001). It has been demonstrated that the level of GA in PSY-overexpressing plants was reduced due to increasing utilization of GGPP for carotenoid biosynthesis, leading to a dwarf phenotype (Fray et al., 1995). In addition, the ratio of ABA/GA is distinctly reduced in phs4-1 seeds, and the genes involved in seed development were differentially expressed in the seeds of the wild type and mutants. These results suggested that the ABA/GA ratio might play an important role in controlling PHS. However, the contribution of carotenoid synthesis to the regulation of balance between ABA and GA is still elusive.

It was found that some common factors regulated ABA and GA biosynthesis during seed development in Arabidopsis in an opposite manner (Gazzarrini et al., 2004). The viviparous mutants of rice are ideal for elucidating the complex mechanism of germination and dormancy, and a detailed comparison on differential expression patterns of genes between different kinds of phs mutants and wild type by microarray or other technologies will finally help us to identify the major factors controlling PHS and gain more insight into the physiological functions of carotenoids and the molecular mechanism of PHS.

Experimental procedures

Plant materials and growth conditions

Approximately 160,000 rice T-DNA/Tos17 mutant lines (Nipponbare background) were sown in the field in Hangzhou, Zhejiang Province in 2003. The panicles of mature plants were observed in late September to detect the germinated seeds. In 2004, the screened phs mutants (T2) were sown in Beijing for further screening. Segregation ratios of viviparous and non-viviparous plants were investigated in the T3 plants. The other two phs3 alleles were obtained as follows: phs3-2 was obtained from TOS17 retrotransposon insertion lines (http://tos.nias.affrc.go.jp/~miyao/pub/tos17/phenotype/17-NG.html, line number NG 0489) and phs3-3 was derived from a spontaneous mutation.

Map-based cloning of PHS genes

To map the mutated genes, linkage analyses were performed using an F2 population derived from the cross between mutants (Nipponbare, Japonica) and TN1 or Minghui 63 (Indica) varieties. The simple-sequence repeat (SSR) and sequence-tagged-site (STS) markers used to analyze the polymorphisms between Nipponbare and TN1 (or Minghui 63) were obtained from the DNA bank at the National Institute of Agrobiological Sciences (http://www.nias.affrc.go.jp).

Sequence analysis

The protein sequences of ABA biosynthetic enzymes from Arabidopsis (http://www.arabidopsis.org/) were compared with the rice database of TIGR Rice Databases (http://www.tigr.org), the Rice Genome Research Program (http://rgp.dna.affrc.go.jp/), the Beijing Genomics Institute (http://bgi.genomics.org.cn/rice), and the Knowledge-based Oryza Molecular biological Encyclopedia (KOME; http://cdna01.dna.affrc.go.jp/cDNA/) for searching the rice orthologs.

Construction of complementation and RNAi vectors and plant transformation

A 6.5-kb DNA fragment containing a full-length genomic β-OsLCY gene was obtained by digesting the bacterial artificial chromosome (BAC) clone OSJNBb0031B09 with PstI and BamHI; the fragment was inserted into a binary vector pCAMBIA2300. Similarly, the complementation vectors for other phs mutants were also constructed, using corresponding BAC clones (Table S3). In addition, empty vectors were transformed into the corresponding phs mutant as the respective controls. For β-OsLCY RNAi construct, an 856-bp fragment from 630 to 1486 bp of the β-OsLCY ORF was inserted as a BamHI/Sall fragment in sense orientation downstream of the potato (Solanum tuberosum L.) GA20 oxidase intron into pUC-RNAi (Luo et al., 2006). The same fragment was inserted in antisense orientation into the BglII/Xhol sites of pUC-RNAi already carrying the sense fragment. Subsequently, the fragment comprising sense and antisense fragments of β-OsLCY interspersed by potato GA20 oxidase intron was excised from pUC-RNAi using the flanking PstI and inserted into a pXQAct plasmid between rice actin1 promoter and Os5 terminator, yielding the binary construct. The rice transformation was performed as described (Liu et al., 2007).

Carotenoid analysis

The extraction and analysis of carotenoids was performed as previously described (Fraser et al., 2000). Briefly, a reverse-phase C30, 5 μm column (250 × 4.6 mm) coupled to a 20 × 4.6 mm C30 guard (YMC Inc.; http://www.ymc.co.jp) with a methanol/tert-methyl butyl ether-based mobile phase were used with a HPLC 10Avp system (Shimadzu, http://www.shimadzu.com/). Throughout chromatography, the elution was monitored continuously from 200 to 600 nm by an online Shimadzu SPD-10Avp PDA detector. Carotenoids were identified by their characteristic absorption spectra, typical retention time, and comparison with authentic standards.

Transmission electron microscopic analysis

Rice leaves were cut into 1 mm squares, fixed in 2.5% (v/v) glutaraldehyde in PBS (pH 7.2) for 24 h and post-fixed in 2% OsO4 in PBS (pH 7.2) for 2 h. Following ethanol series dehydration, samples were embedded in Epon 812 (Shell Chemicals; http://www.shellchemicals.com) and polymerized for 24 h at 60°C. Ultrathin sections (50–70 nm) were double stained with uranyl acetate and lead citrate and examined with a transmission electron microscope (TEM; FEI Tecnai 20; http://www.fei.com) at 120 kV.

Histochemical detection of O2-

Superoxide accumulation in rice leaves was visualized by 0.1% NBT staining as described (Fitzgerald et al., 2004). Rice leaves were...
Chlorophyll fluorescence measurements

Chlorophyll fluorescence measurements were carried out with attached leaves in the greenhouse using a PAM 2100 portable fluorometer (Walz, http://www.walz.com/) as described (Lu et al., 2001).

Determination of ABA, GA, and water-loss assay

Determination of ABA and GA with 2-week-old seedlings was performed as previously described (Agrawal et al., 2001; Luo et al., 2006). For the water-loss assay, plants were grown in a paddy field and the detached leaves (300 mg from 10 seedlings) of 1-month-old seedlings were kept on aluminum foil at room temperature (25°C). The weight of leaves was taken at 10-min intervals until 60 min or longer. Each experiment was performed with five replicates.

Germination and ABA response assay

Seeds (30 days after pollination) from phs mutants, the wild type and complementation transgenic plants were sampled for germination ratio and ABA response assay. Seeds were cultured in a growth chamber at 30°C/C with a 16-h/8-h cycle for 3 days for the germination assay and 11 days for the ABA response assay. Shoots from seeds without ABA treatment were measured regularly, while shoots from seeds treated with different concentrations of ABA were measured at the 11th day after growth.

Expression analysis

Total RNAs were isolated from various organs of rice as described (Luo et al., 2006). The RT-PCR was performed with DNase-treated total RNAs using the RT-for-PCR Kit (Promega, http://www.promega.com/). The primers used for each gene are listed in Table S1. The products of PCR amplification were separated by electrophoresis on 1.5% (w/v) agarose gels and stained with ethidium bromide.

Western blot analysis

Thylakoid were extracted from the first leaves of β-OsLCY-RNAi plants, phs3-1 mutant and the wild type when β-OsLCY-RNAi plants and phs3-1 mutant showed a photobleaching phenotype at the tillering stage. Thylakoid membrane preparation and immunodetection of the PS II core proteins CP43, CP47 and D1 were carried out as described previously (Chen and Gallie, 2004; Gonzalez et al., 1998; Jiang and Zhang, 2002; Knörrer et al., 1996).

Enzyme activity assay

All enzyme activities were measured from 0.5 g first leaves of β-OsLCY-RNAi plants and the phs3-1 mutant as well as the wild type when β-OsLCY-RNAi plants and the phs3-1 mutant showed a photobleaching phenotype at the tillering stage. The enzyme activities of SOD, CAT, APX, MDAR, DHAR, and GR were measured as described previously (Chen and Gallie, 2004; Gonzalez et al., 1998; Jiang and Zhang, 2002; Knörrer et al., 1996).

Pre-harvest sprouting in rice

We are grateful to Dr. Jianru Zuo of IGDB, CAS for critically reading the manuscript. This work was supported by grants from the Chinese Academy of Sciences (KSCX2-YW-N-010) and Ministry of Science and Technology of China (2002CCA03200, 2002CB111301, and 2005CB120806) and also the National Natural Sciences Foundation of China (3062100, 30425034, 3071013903).

Supplementary Material

The following supplementary material is available for this article online:

Figure S1. Carotenoid and abscisic acid biosynthetic pathway.

Figure S2. The phenotypes of the rice pre-harvest sprouting mutants.

Figure S3. Activities of ROS-scavenging enzymes of leaves from OsLCY-RNAi plant, phs3-1, and wild type at tillering stage.

Figure S4. Abscisic acid responsiveness test of phs3-1 mutant.

Figure S5. Map-based cloning of PHS genes.

Table S1. The primers used for expression analysis via RT-PCR.

Table S2. Segregation ratios of viviparous and non-viviparous plants in the T2 generation.

Table S3. Constructions for genetic complementation of phs mutants and plant transformation events.

This material is available as part of the online article from http://www.blackwell-synergy.com. Please note: Blackwell publishing are not responsible for the content or functionality of any supplementary materials supplied by the authors. Any queries (other than missing material) should be directed to the corresponding author for the article.

References


Fitzgerald, H.A., Chern, M.S., Navarre, R. and Ronald, P.C. (2004) Overexpression of (At)NPR1 in rice leads to a BTH- and environ-


Murashige, T. and Skoog, F. (1962) A revised medium for rapid
growth and bio assays with tobacco tissue cultures. Physiol.
Plant. 15, 473–497.


mutants define a central role for the xanthophyll cycle in the
regulation of photosynthetic energy conversion. Plant Cell, 10,
1121–1134.

Niyogi, K.K., Shih, C., Soon Chow, W., Pogson, B.J., Dellapenna, D.
and Bjorkman, O. (2001) Photoprotection in a zeaxanthin- and
67, 139–145.

O’Neill, S.D., Kumagai, M.H., Majumdar, A., Huang, N., Sutliff, T.D.
sativa: characterization of cDNA clones and mRNA expression

promoter of rice contains two distinct abscisic acid-responsive

(2002) Identification of the carotenoid isomerase provides
insight into carotenoid biosynthesis, prolamellar body formation,

Peng, L., Ma, J., Chi, W., Guo, J., Zhu, S., Lu, Q., Lu, C. and Zhang, L.
(2006) LOW PSII ACCUMULATION1 is involved in efficient
assembly of photosystem II in Arabidopsis thaliana. Plant Cell, 18,
955–969.

Prada, D., Ullrich, S.E., Molina-Cano, J.L., Cistue, L., Clancy, J.A. and

and Boronat, A. (2001) 1-Deoxy-D-xylulose 5-phosphate reducto-
isomerase and plastid isoprenoid biosynthesis during tomato
fruit ripening. Plant J. 27, 213–222.

Sagar, A.D., Horwitz, B.A., Elliott, R.C., Thompson, W.F. and Briggs,
W.R. (1988) Light effects on several chloroplast components in

golden rice golden (yellow) instead of red? Plant Physiol. 138,
441–450.

indirect pathway of abscisic acid biosynthesis by mutants, genes,

Singh, M., Lewis, P.E., Hardeman, K., Bai, L., Rose, J.K., Mazurek,
M., Chomet, P. and Brutnell, T.P. (2003) Activator mutagenesis of
the pink scutellum1/viviparous7 locus of maize. Plant Cell, 15,
874–884.

mapping enables dissection of closely linked quantitative trait
107, 1174–1180.

Tracewell, C.A., Vrettos, J.S., Bautista, J.A., Frank, H.A. and Brudvig,

Zhang, L.X. (2006) Genetic dissection of the seed dormancy trait
in cultivated rice (Oryza sativa L.). Plant Sci. 170, 786–792.

Welsch, R., Beyer, P., Hugueney, P., Kleing, H. and von Lintig, J.
(2000) Regulation and activation of phytoene synthase, a key
enzyme in carotenoid biosynthesis, during photomorphogenesis.
Planta, 211, 846–854.

White, C.N. and Rivin, C.J. (2000) Gibberellins and seed develop-
ment in maize. II. Gibberellin synthesis inhibition enhances
abscisic acid signaling in cultured embryos. Plant Physiol. 122,
1089–1097.

Gibberellins and seed development in maize. I. Evidence that
gibberellin/abscisic acid balance governs germination versus

Wilkinson, M.D., Mckibbin, R.S., Bailey, P.C. and Flintlham, J.E.
(2002) Use of comparative molecular genetics to study preharvest
sprouting in wheat. Euphytica, 126, 27–33.

Woodall, A.A., Britton, G. and Jackson, M.J. (1997) Carotenoids and
protection of phospholipids in solution or in liposomes against
oxidation by peroxyl radicals: relationship between carotenoid
structure and protective ability. Biochim. Biophys. Acta, 1336,
575–586.

identify the first rice mutants blocked in carotenoid biosynthesis.
J. Exp. Bot. 52, 161–166.

Xiong, L. and Zhu, J.K. (2003) Regulation of abscisic acid biosyn-