



## Effect of metal ion on the structure and function of LiPDF: The study of the fine structure around the metal site using XANES

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### ABSTRACT

We used X-ray absorption near edge structure (XANES) spectroscopy to investigate the metal-dependent enzymatic activity of the peptide deformylase from *Leptospira interrogans* (LiPDF). *Ab initio* full multiple scattering calculations performed by MXAN are applied to obtain the local structure of the cobalt-containing LiPDF (Co-LiPDF) and zinc-containing LiPDF (Zn-LiPDF) around the metal sites in pH9.0 buffer solution. The result shows the cobalt-wat1 (the bond water molecule) distance of Co-LiPDF is 1.89 Å, much shorter than that of Zn-LiPDF, 2.50 Å. That is an essential factor for its low catalytic activity.

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### 1. Introduction

Peptide deformylase (PDF, EC 3.5.1.27) is an important member of the zinc metalloproteases superfamily. In eubacterium, as well as in mitochondria and chloroplasts, the protein synthesis starts from formylmethionine-tRNA<sub>f</sub>, resulting in the synthesis of N-terminally formylated polypeptides [1]. During the elongation of the polypeptide chain the formyl group is removed hydrolytically by the PDF enzyme [2–4], which may be a universally conserved feature of the prokaryotic protein synthesis and an essential component for bacterial survival [5]. Actually there is an increasing demand from new effective drugs because of the continuous increase of tolerance problems occurring with conventional antibiotics. PDF is an attractive target for the design of new antibiotics because deformylation apparently does not occur in the cytoplasm of eucaryotic cells [6].

Previous work has shown that the catalytic efficiency of PDFs strongly depend on the identity of the bound metal. *Escherichia coli* PDF (EcPDF) containing Zn<sup>2+</sup> is much less active than that containing Fe<sup>2+</sup>, Ni<sup>2+</sup> and Co<sup>2+</sup>, while for *Leptospira interrogans* PDF (LiPDF), that containing Zn<sup>2+</sup> is more active than others [7–9]. Little is known about the chemical and physical mechanisms responsible for the enzymatic activity difference of LiPDFs due to the lack of the fine structure around the metal sites in these proteins.

X-ray absorption spectroscopy (XAS) is a unique tool to probe the local structure of metalloproteins around metal sites at atomic-level resolution [10–15], where biological samples can be studied in a variety of forms, including oriented single protein crystals, unoriented microcrystals, frozen or room-temperature solutions and proteins inserted in membranes. Here, we measured the Zn and Co K-edge XAS spectra for zinc-containing LiPDF (Zn-LiPDF) and cobalt-containing LiPDF (Co-LiPDF), respectively, in pH 9.0 buffer solution. Combining *ab initio* multiple scattering calculation, we extracted the subtle local structure information around metal sites for both LiPDF proteins, especially the position of the bond water molecule.

### 2. Materials and methods

*E. coli* BL21(DE3) cells carrying plasmid pET22b-LiPDF were expressed as previously reported [7]. Cells were harvested by centrifugation, and resuspended in 25 ml of buffer A (50 mM Tris-HCl 7.5, 10 mM NaCl). After sonication cell debris was removed by centrifugation. The supernatant (25 ml) was applied onto a HiTrapTM 5 ml Q HP (Amersham Pharmacia Biotech) column equilibrated with buffer B (50 mM Tris-HCl pH 8.5, 10 mM NaCl). The column was eluted with 250 ml of buffer B plus a linear gradient of 10 mM–1 M NaCl (AKTA purifier 900, Amersham Pharmacia Biotech). The fractions with PDF activity were concentrated and applied to a SuperdexTM 75 column (16/60, Amersham Pharmacia Biotech) pre-equilibrated with buffer C (50 mM Tris-HCl pH 8.5, 50 mM NaCl). The fractions containing LiPDF activity were collected. The sample homogeneity was monitored by SDS-PAGE and the protein concentration was determined by Bradford assay with BSA as the standard. Finally,

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the purified protein was dialyzed against 50 mM  $\text{NaH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$  (pH 9.0) and concentrated to about 60 mg/ml. The  $\text{Zn}^{2+}$  concentration is about 200 ppm. The enzymatic activity of Zn-*LiPDF* is nearly constant from pH 7.5–10.5 [7]. We choose pH 9.0 just as it is in the middle of the active range.

Samples of Co-*LiPDF* were prepared by incubation of metal-free *LiPDF*. Metal-free *LiPDF* was prepared by adding 15 ml of 0.1 M EDTA to 0.5 ml of 60 mg/ml protein for 2 hours at 277 K. There was no Zn element left as monitored by X-ray fluorescence method. The additional EDTA was then removed by direct exchange dialysis and concentrated in centrifugal tube. The procedure was repeated twice by direct exchange dialysis (dilute  $100 \times 100$  times) and concentrated in centrifugal tube (dilute  $15 \times 15$  times). Metal-free *LiPDF* (0.5 ml, 20 mg/ml) was incubated with 1.2 mM  $\text{CoCl}_2$  60 ml for 2 hours at 277 K and then concentrated to 20 mg/ml. The purified Co-*LiPDF* was dialyzed against 50 mM  $\text{NaH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$  (pH 9.0) and afterwards concentrated to 30 mg/ml. The  $\text{Co}^{2+}$  concentration is about 100 ppm. The sample homogeneity was monitored by SDS-PAGE. The protein concentration was finally determined by Bradford assay with BSA as the standard [16].

Measurements of the Zn *K*-edge and Co *K*-edge were carried out at the beam line 4W1B of the Beijing Synchrotron Radiation Facility (BSRF, China) in the fluorescence yield (FY) mode at room temperature. The typical energy of the storage ring was 2.5 GeV and experiments were performed with a decreasing electron current from 250 to 160 mA. The incident beam intensity was monitored using an ionization chamber flooded by a 25% argon-doped nitrogen mixture while the fluorescence signal was collected by means of a Lytle detector flooded by argon gas.

In order to extract the structural/geometrical information around the metal site, we performed a quantitative analysis of the XANES spectrum using the MXAN code [17,18]. This code is capable of performing a quantitative analysis of a XANES spectrum from the absorption edge up to 200 eV via comparison between experimental data and many theoretical calculations obtained by changing relevant geometrical parameters of the atomic site [19–21]. The X-ray absorption cross-sections are calculated using the full multiple scattering scheme in the framework of the muffin-tin (MT) approximation for the shape of the potential [22,23]. In this case, the exchange and correlation part of the potential have been determined on the basis of the local density approximation of the self energy. Inelastic processes have been taken into account by a convolution with a broadening Lorentzian function having an energy-dependent width of the form  $\Gamma(E) = \Gamma_c + \Gamma_{\text{mfp}}(E)$ . The constant part  $\Gamma_c$  includes the core-hole lifetime (1.8 eV) [24] and the experimental resolution (1.5 eV), while the energy-dependent term represents all the intrinsic and extrinsic inelastic processes [21,25,26]. The muffin-tin radii were chosen according to the Nofrman criterion with a 15% overlap [27,28]. This method takes into account, for Multiple-Scattering events, a rigorous way through the evaluation of the scattering path operator [22,29] and its effectiveness has been successfully tested over the years in a number of different applications [19,25]. The spectrum is convergent when the cluster includes 39 atoms within a radius of 5.5 Å. Therefore the local structure of the *LiPDF* active center can be properly represented by a cluster that contains all the 50 atoms within 6 Å from zinc ion.

### 3. Results and discussion

The Zn *K*-edge XANES spectrum of Zn-*LiPDF* (red curve) and the Co *K*-edge XANES spectrum of Co-*LiPDF* in buffer solution at pH 9.0 are shown in Fig. 1. Here the zero energy corresponds to

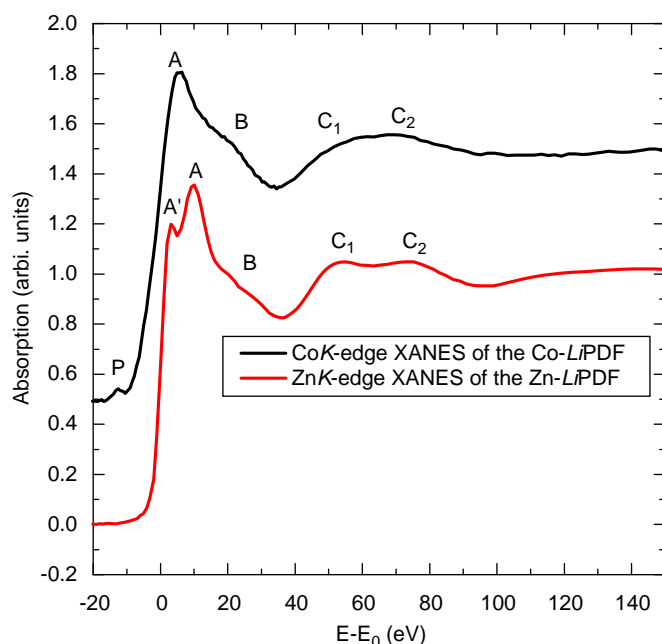
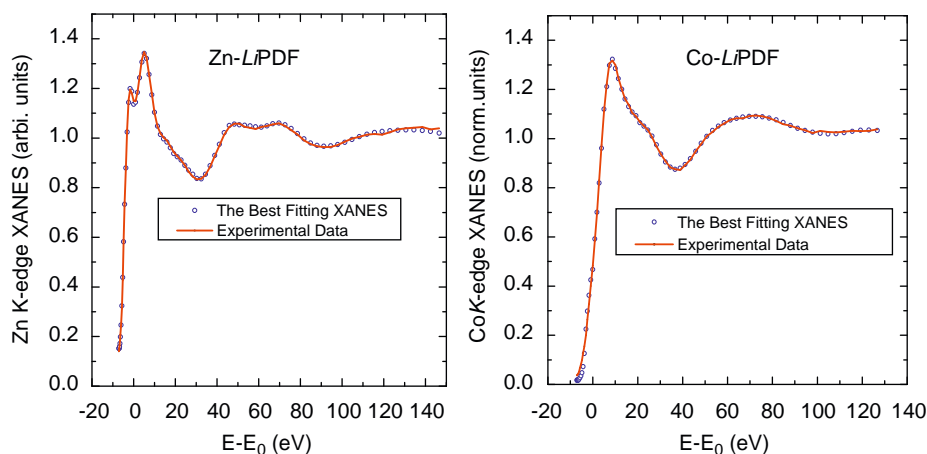


Fig. 1. Zn *K*-edge XANES spectrum of Zn-*LiPDF* (red curve) and Co *K*-edge XANES spectrum of Co-*LiPDF* in buffer solution at pH 9.0. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

the absorption edge of the two spectra. It can be seen that both of the XANES spectra present the similar features above 20 eV. As we know, the XAS feature of this region is mainly dominated by the scattering events of the neighbor atoms around the absorbed atom, but not sensitive to the electronic structure of metals. It indicates  $\text{Co}^{2+}$  and  $\text{Zn}^{2+}$  locate the similar chemical environment in the *LiPDF* proteins. However, the subtle change of the two XANES spectra also exists, especially for the peaks  $C_1$  and  $C_2$ , indicating the fine local structure changes when  $\text{Co}^{2+}$  substitutes  $\text{Zn}^{2+}$ . Here it should be noted that the big difference present in the region  $-10$  to  $20$  eV is due to the different 3d electronic configuration of metals. An obvious pre-edge peak can be observed in the XANES spectrum of the Co-*LiPDF*, because of the unoccupied 3d orbital. The peak A is due to the scattering among the nearest neighbors and it can reflect not only the geometric structure but also the electronic structure. Due to the electronic configure of the  $\text{Zn}^{2+}$  is different from that of  $\text{Co}^{2+}$ , so we can not directly compare the main peak A in the spectrum of Co-*LiPDF* with that of Zn-*LiPDF*.

In order to obtain the exact structural information around the metal site, we performed a quantitative analysis of the XANES spectrum using the MXAN code, based on the crystallographic structure of *LiPDF* (PDB code 1Y6H(B)) [30,31]. Fig. 2 shows the best fits (blue dots) of the experimental XANES spectra (red curve) of Zn-*LiPDF* (left) and Co-*LiPDF* (right) in buffer solution at pH 9.0. As we can see, all features in the near-edge region of the XANES spectra at the Zn and Co *K*-edge have been successfully reproduced. The square residue values for these calculations are 0.45 and 0.87, respectively. Some refined critical distances of the best fits are summarized in Table 1. The comparison of the local structure obtained by the best fits for the Zn-*LiPDF* and the Co-*LiPDF* is plotted in Fig. 3.

Our result shows the most important difference of the local structure around center metal between Co-*LiPDF* and Zn-*LiPDF* is the position of the wat1. The cobalt-wat1 distance of Co-*LiPDF* is 1.89 Å, much shorter than that of Zn-*LiPDF*, 2.50 Å. In most of PDF proteins and PDF complexes, the length of the metal-wat1 bond

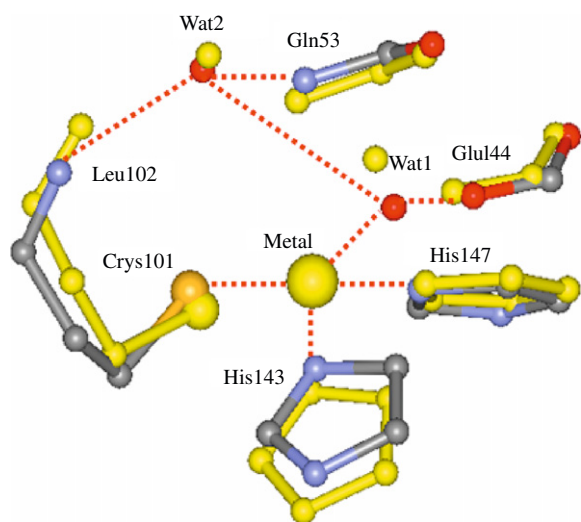


**Fig. 2.** Best fit of the XANES spectra (blue dots) of Zn-LiPDF (left) and Co-LiPDF (right) in buffer solution at pH9.0, compared with experimental spectra (red line). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

**Table 1**

Refined critical distances of Zn-LiPDF and Co-LiPDF.

Atom pairs	Model structure distance (Å)	Zn-LiPDF fitting distance (Å)	Co-LiPDF fitting distance (Å)
S <sup>2</sup> (square residue)	–	0.45	0.87
Metal His147-(NE2)	2.15	2.03 ± 0.02	2.03 ± 0.03
Metal His143-(NE2)	2.17	2.04 ± 0.02	1.85 ± 0.02
Metal Cys101-(SG)	2.40	2.23 ± 0.02	2.32 ± 0.03
Metal Wat1-(OH2)	2.31	2.50 ± 0.03	1.89 ± 0.03
Metal Wat2-(OH2)	3.90	4.00 ± 0.04	3.59 ± 0.04
Wat1-(OH2) Wat2-(OH2)	3.20	2.95 ± 0.05	3.53 ± 0.05
Wat1-(OH2) Glu144-(OE2)	2.67	2.48 ± 0.06	3.00 ± 0.07



**Fig. 3.** Comparison between the local structure of the best fit of Co-LiPDF (colored sticks) and Zn-LiPDF (thick yellow sticks) at pH9.0. The coordination of the metal ion is indicated in red dash line. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

varies between 1.80 and 2.63 Å. So we think the length of the Co-wat1 bond and Zn-wat1 bond in our measurement are reasonable. This stronger water-bonding of cobalt inhibits the transition from the tetrahedral to a five-coordinate metal center in the catalytic process and makes the distance between water and Glu144 (wat1-Glu144) become longer. In the catalytic process of PDF

proteins, the very important step is that the proton of wat1 transfer to the amide at the N-terminus of the peptide with the help of Glu144 and the added positive charge makes the nitrogen suitable as a leaving group [9]. Therefore, lengthening of the wat1-Glu144 distance weakens the polarization ability of Glu144 to wat1, resulting in the proton of wat1 hardly transfer into the amide at the N-terminus.

#### 4. Conclusion

In this work we have investigated metal-dependent enzymatic activity of the peptide deformylase from *Leptospira interrogans* (PDB code 1Y6H) by using XANES spectroscopy combining with *ab initio* multiple scattering calculations. All features in the near-edge region of the XAS spectra at the Zn and Co K-edge have been successfully reproduced and detailed structural information around the metal ion has been obtained. The result shows that cobalt ion presents the stronger water-bonding than zinc ion in LiPDF proteins. It weakens the polarization ability of Glu144 to wat1, resulting in the proton of wat1 hardly transfer to the amide at the N-terminus. It is the reason that the enzymatic activity of Co-LiPDF is much lower than that of Zn-LiPDF.

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